

Relationship Between ANCA-Associated Vasculitis and Infection: A Review

Yingxue Zhou, Qing Shen

Department of Nephrology, The First Affiliated Hospital of Chongqing Medical University, Chongqing 400010, China.

Abstract: ANCA-associated vasculitis (AAV) is an autoimmune disease with multiorgan involvement of the entire body. And infection has been paid more and more attention in the course of treatment. As far as the current research is concerned, infection participates in the pathogenesis of AAV, interferes with the treatment of AAV, and affects the prognosis of AAV. This paper focuses on the role of pathogen infections in AAV pathogenesis. This review also elaborates on the types and prognosis of secondary infections in AAV patients. According to the current study, maintaining an appropriate BMI and vaccination is beneficial to the prevention of infection and the prognosis of AAV patients.

Keywords: ANCA; Infection; Pathogenesis; Complications.

Introduction

Antineutrophil Cytoplasmic Antibody (ANCA) associated vasculitis (AAV) is characterized by inflammation of small vessels. It is an autoimmune disease that can affect a number of different systems of the body. The ANCA serotypes are PR3-ANCA, MPO-ANCA, and ANCA-negative. According to pathological classification, it is divided into Microscopic polyangiitis (MPA), Granulomatosis with polyangiitis (GPA), and Eosinophilic granulomatosis with polyangiitis (EGPA).^[1] Although AAV is rare, the occurrence rate of AAV has been increasing over time.^[2] And infection is involved in all stages of AAV. This review aims to illustrate the relationship between AAV and infection.

1. Pathogenesis

ANCA is an autoantibody that is in the neutrophils and monocytes. A case in 2009 reported that MPO-ANCA from a mother with MPA passed through the placenta into a preterm infant. Subsequently, the infant developed pulmonary hemorrhage and renal damage in a few days.^[3] This report provides direct evidence that ANCA is pathogenic in humans. The main ANCA target antigens are Proteinase 3 (PR3) and Myeloperox-idase (MPO). Neutrophils and monocytes can be activated by MPO-ANCA and PR3-ANCA. Activated neutrophils undergo respiratory bursts and degranulation.^[4] Similar responses were also observed in monocytes. ^[5] And then a series of inflammatory reactions happen and finally lead to vasculitis.

There are currently two hypotheses:

1) Molecular mimicry hypothesis: In a study, anti-human Lysosome-associated membrane protein 2 (LAMP-2) autoantibodies have also been found to participate in the pathogenesis of ANCA. There is 100% homology between bacterial adhesins FimH and LAMP-2, so FimH is considered to trigger autoimmunity to LAMP-2.^[6]

2) Autoantigen complementation hypothesis: A study found antibodies against the complementary peptide of PR3 (cPR3) in some PR3-ANCA-positive patients.^[7] Therefore, the initial immune response in AAV patients is not against the

self-antigen, but against the peptide complementary to the self-antigen epitope. In recent years, it has been found that neutrophils, Neutrophilextracellular traps (NETs), complement, and Lymphocytes all play a big part in AAV pathogenesis.

2. Infection induces AAV

According to the above two hypotheses, infectious factors are involved. The first hypothesis suggests that FimH is present in some Gram-negative bacteria. When humans are infected with FimH-bearing pathogens, it may lead to AAV. From the second hypothesis, pathogens with constitutive analogues of cPR3, such as Staphylococcus aureus and Ross River virus, could presumably act as exogenously introduced cPR3 to cause ANCA formation.

2.1 Staphylococcus aureus

In Stegeman's study, nasal S. aureus was present in 63% of patients with GPA in the experimental group. It was 25% in the control group. Meanwhile, there was a remarkably increased risk of GPA recurrence in the experimental one. [8] A 2017 study similarly confirmed that GPA patients with active nasal disease at admission were more likely to recur.^[9] It has been demonstrated that superantigens, peptidoglycans, and fungal β -glucans derived from Staphylococcus aureus can induce the expansion of Th17 cells. ^[10] Th17 cells are considered central parts of the autoimmune response. ^[11] In addition, Staphylococcus aureus can induce NETs formation.^[12] NETs destroy vascular endothelial cells. And they activate Lymphocytes to promote autoimmune responses.^[13]

2.2 Viruses

It has been proposed that cytomegalovirus, HBV, HCV, etc., participate in the pathogenesis of autoimmune diseases.^[14] Many studies have confirmed that SARS-CoV-2 can cause autoimmune diseases. The mechanism by which COVID-19 induces AAV is not clear. A theory suggests that COVID-19 could lead to an increase in proinflammatory cytokines and cytokine storm.^[15] Another hypothesis is that COVID-19 causes transient immunosuppression and inappropriate immune reconstitution, causing the development of autoantibodies and the inability to correctly recognize self-antigens.^[16]

3. Secondary infection

Immunosuppressive therapy can effectively improve the survival time and quality of life of AAV patients. But increased infection morbidity and mortality were found in AAV patients during immunosuppressive therapy. In a recent study, the cumulative rate of infection in AAV patients at 1 and 5 years was almost 50% and 65%. Serious infections were almost 20% and 25%. Respiratory tract infections were the most common. And Staphylococcus aureus was the highest proportion of pathogen cultures (41%).^[17]

Another study found that the highest incidence of serious infection in AAV patients at 1 and 5 years was almost 20% and 10%, both lower than the rates in those studies. The study attributes this reduction to better long-term care, controlled long-term use of CYC, and limited initial prednisone use.^[18] The 2021 KDIGO guidelines state that initial therapy for de novo AAV can be induction therapy with CYC or RTX plus glucocorticoids.^[19] The happening of serious infections was found to be more in patients receiving CYC (20%) than RTX (11%) during the total induction treatment. The respiratory tract infections were 45%, followed by 24% herpes zoster.^[18]

A Chinese study also found that lung infection was the most common type of infection (72%). Pathogens found during infection include bacteria, fungi, and viruses. the most common infections were bacterial (65%), particularly Acinetobacter baumannii and Staphylococcus aureus, followed by fungal (25%) and viral (10%) infections. These include cytomegalovirus and Pneumocystis.^[20]

4. Prevention and prognosis

One study found that 14% of AAV patients who were infected during treatment died of severe pneumonia after immunosuppressive therapy.^[18] In a large sample study in China, the cumulative survival rates at 1 and 5 years were almost 80% and 60%. Secondary infections were the major cause of death in the first year after diagnosis (40%).^[21]

Kinds of literature has suggested that we should pay attention to the dose of immunosuppressive agents and the speed of dose reduction to avoid infection. For AAV patients treated with CTX or RTX ,trimethoprim-sulfamethoxazole is advised for the prophylaxis of Pneumocystis infection.^[22] Guidelines recommend that most AAV patients receive vaccines, such as the influenza vaccine, pneumonia vaccine, and others.^[23] A Japanese study found that malnutrition in patients at the time of AAV diagnosis may increase vulnerability to infection during immunosuppressive therapy.^[24] Therefore, it is necessary to maintain an appropriate BMI in dealing with infection.

This review preliminarily elucidates the role of infection in the pathogenesis, progress, and prognosis of AAV. Now there are still no clear guidelines and norms for the treatment of AAV complicated with infection. There are still contradictions between immunosuppressive therapy and anti-infection therapy. Doctors can only make corresponding judgments according to the patient's individual condition and give the patient appropriate treatment measures.

References

[1] Jennette J, Falk R, Bacon P, *et al.* 2012 Revised International Chapel Hill Consensus Conference Nomenclature of Vasculitides[J]. Arthritis & Rheumatology, 2012, 20(1).

[2] Mohammad AJ. An update on the epidemiology of ANCA-associated vasculitis. Rheumatology (Oxford). 2020 May 1;59(Suppl 3):iii42-iii50.

[3] Silva F, Specks U, Sethi S, *et al.* Successful Pregnancy and Delivery of a Healthy Newborn Despite Transplacental Transfer of Antimyeloperoxidase Antibodies From a Mother With Microscopic Polyangiitis[J]. American Journal of Kidney Diseases, 2009, 54(3):542-545.

[4] Falk RJ, Terrell RS, Jennette L, *et al.* Anti-Neutrophil Cytoplasmic Autoantibodies Induce Neutrophils to Degranulate and Produce Oxygen Radicals in vitro[J]. Proceedings of the National Academy of Sciences of the United States of America, 1990, 87(11):4115-4119.

[5] Jennette JC, Falk RJ. ANCAs Are Also Antimonocyte Cytoplasmic Autoantibodies[J]. Clinical Journal of the American Society of Nephrology Cjasn, 2014, 10(1):4-6.

[6] Kain R, Exner M, Brandes R, *et al.* Molecular mimicry in pauci-immune focal necrotizing glomerulonephritis[J]. Nature Medicine, 2008, 14(10): 1088-1096.

[7] Pendergraft WF, Preston GA, Shah RR, *et al.* Autoimmunity is triggered by cPR-3(105-201), a protein complementary to human autoantigen proteinase-3.[J]. Nature Medicine, 2004, 10(1):72-79.

[8] Stegeman CA, Tervaert JW, Sluiter WJ, *et al.* Association of Chronic Nasal Carriage of Staphylococcus aureus and Higher Relapse Rates in Wegener Granulomatosis[J]. Ann Int Med, 1994, 120(1):12-17.

[9] Salmela A, Rasmussen N, Tervaert JWC, *et al.* European Vasculitis Study Group. Chronic nasal Staphylococcus aureus carriage identifies a subset of newly diagnosed granulomatosis with polyangiitis patients with high relapse rate. *Rheumatology (Oxford)*. 2017;56(6):965-972.

[10] Li H, Nooh MM, Kotb M, *et al.* Commercial peptidoglycan preparations are contaminated with superantigen-like activity that stimulates IL-17 production[J]. Journal of Leukocyte Biology, 2008, 83(2): 409-418.

[11] Oukka, M. Th17 cells in immunity and autoimmunity[J]. Annals of the Rheumatic Diseases, 2008, 67 Suppl 3(Suppl 3):986789-986789.

[12] Bhattacharya M, Berends ETM, Chan R, et al. Staphylococcus aureus biofilms release leukocidins to elicit extracellular trap formation and evade neutrophil-mediated killing. Proc Natl Acad Sci U S A. 2018;115(28):7416-7421.

[13] Nakazawa D, Masuda S, Tomaru U, et al. Pathogenesis and therapeutic interventions for ANCA-associated vasculitis [published correction appears in Nat Rev Rheumatol. 2019 Jan 17;:]. Nat Rev Rheumatol. 2019;15(2):91-101.

[14] Sfriso P, Ghirardello A, Botsios C, *et al.* Infections and autoimmunity: the multifaceted relationship. J Leukoc Biol. 2010;87(3):385-395.

[15] Henderson LA, Canna SW, Schulert GS, *et al.* On the alert for cytokine storm: Immunopathology in COVID-19[J]. Arthritis & Rheumatology, 2020, 72(7).

[16] Caas CA . The triggering of post-COVID-19 autoimmunity phenomena could be associated with both transient immunosuppression and an inappropriate form of immune reconstitution in susceptible individuals[J]. Medical Hypotheses, 2020, 145:110345.

[17] Mcgregor J, Negrete-Lopez R, Poulton C J, *et al.* Adverse events and infectious burden, microbes and temporal outline from immunosuppressive therapy in antineutrophil cytoplasmic antibody-associated vasculitis with native renal function[J]. Nephrology Dialysis Transplantation, 2015, 30(suppl 1): i171-i181.

[18] Thomas K, Argyriou E, Kapsala N, *et al.* Serious infections in ANCA-associated vasculitides in the biologic era: real-life data from a multicenter cohort of 162 patients[J]. Arthritis Research & Therapy, 2021, 23(1):1-9.

[19] Kidney Disease: Improving Global Outcomes (KDIGO) Blood Pressure Work Group. KDIGO 2021 Clinical Practice Guideline for the Management of Blood Pressure in Chronic Kidney Disease. Kidney Int. 2021 Mar;99(3S):S1-S87.

[20] Yang L, Xie H, Liu Z, et al. Risk factors for infectious complications of ANCA-associated vasculitis: a cohort study[J]. Bmc Nephrology, 2018, 19(1):1-7.

[21] Lai QY, Ma TT, Li ZY, *et al.* Predictors for mortality in patients with antineutrophil cytoplasmic autoantibody-associated vasculitis: a study of 398 Chinese patients.[J]. Journal of Rheumatology, 2014, 41(9):1849-55.

[22] Kronbichler A, Kerschbaum J, Gopaluni S, *et al.* Trimethoprim-sulfamethoxazole prophylaxis prevents severe/life-threatening infections following rituximab in antineutrophil cytoplasm antibody-associated vasculitis. Ann Rheum Dis. 2018;77(10):1440-1447.

[23] Furer V, Rondaan C, Heijstek MW, et al. 2019 update of EULAR recommendations for vaccination in adult patients with autoimmune inflammatory rheumatic diseases. Ann Rheum Dis. 2020;79(1):39-52.

[24] Sugiyama H, Yamaguchi M, Katsuno T, *et al.* Association between body mass index and severe infection in older adults with microscopic polyangiitis: a retrospective cohort in Japan[J]. BMC Geriatrics, 2021, 21(1):1-10.